

# Ult raSYBR Mixture (Low ROX)

Project number: U665694

Storage condition:  $-20^{\circ}$ C, if need to use frequently, can be stored in  $2-8^{\circ}$ C,

try to avoid repeated freezing and thawing.

# Product content

Component	U665694-5ml	U665694-40m1
2 x UltraSYBR Mixture (Low ROX)	$5 \times 1m1$	40 x 1m1
ddH2O	$5 \times 1m1$	40 x 1m1

#### Product Introduction

UltraSYBR Mixture (Low ROX) is a premixed system dedicated to dye-based (SYBR Green I) real-time fluorescent quantitative PCR at a concentration of  $2\times$ , containing GoldStar Taq DNA Polymerase, PCR Buffer, dNTPs, SYBR Green I fluorescent dye and Mg2+ and Low ROX correction dyes, which is easy and convenient to operate. It is mainly used for the detection of genomic DNA target sequences and cDNA target sequences after RNA reverse transcription. The fluorescent dye SYBR Green I contained in this product can bind to all double-stranded DNAs, enabling the product to be used for the detection of different target sequences without the need to synthesize specific labeling probes. GoldStar Taq DNA Poly-merase is a chemically modified, new and highly efficient hot-start enzyme, which has no polymerase activity at room temperature, effectively avoiding non-specific amplification caused by nonspecific binding of primers and templates or primer dimerization at room temperature, and the enzyme activation must be incubated at 95℃ for 10 minutes. The unique combination of PCR buffer system and hot start enzyme effectively inhibits non-specific PCR amplification and significantly improves PCR amplification efficiency.

The ROX dye contained in this kit can correct the fluorescence signal error generated between the wells of the quantitative PCR instrument. The low content of ROX correction dye in this kit is suitable for ABI Prism7500/7500 Fast, Stratagene Mx3000/Mx3005P, Corbett Rotor Gene 3000 and other fluorescence quantitative PCR instruments that require a low ROX signal correction. for ABI Prism7500/7500 Fast, Stratagene Mx3000/ Mx3005P, Corbett Rotor Gene 3000, etc. that require lower ROX signal correction.

## Product Features

1. The new high-efficiency hot-start enzyme GoldStar Taq DNA Polymerase and unique PCR buffer system are used in this product, which significantly improves the amplification efficiency of PCR with high sensitivity and specificity.

2. Suitable for fluorescence quantitative PCR assay, which can accurately

quantify and detect the target gene.

### matters needing attention

- 1. Before use, please mix it gently by turning it up and down, avoid foaming as much as possible, and use it after centrifugation for a short time.
- 2. This product contains SYBR GreenI fluorescent dye and ROX dye, avoid strong light when storing this product or preparing PCR reaction solution.
- 3. Avoid repeated freezing and thawing of the product, repeated freezing and thawing may degrade the performance of the product.



- 4. This product cannot be used for fluorescence quantitative PCR by probe method.
- 5. When preparing the reaction solution, please use new or non-contaminated tips and centrifuge tubes to prevent contamination as much as possible.

## Usage

The following examples are conventional PCR reaction systems and reaction conditions, which should be improved and optimized according to the template, primer structure and target fragment size in actual operation.

1. PCR reaction system

reagents	50μl reaction system	final concentration
2×UltraSYBR Mixture(Low ROX)	25 μ1	1×
Forward Primer, 10µM	1 μ 1	0. 2 μ M <sup>1</sup> )
Reverse Primer, 10µM	1 μ 1	0.2 μ M <sup>1</sup> )
Template DNA	2 μ 1²)	
ddH <sub>2</sub> 0	up to 50 µ 1	

Note: 1) Usually, a primer concentration of 0.2  $\mu$ M gives better results, and a final concentration of 0.1-1.0  $\mu$ M can be used as a reference for setting the range.

- (2) Usually the amount of DNA template is 10-100ng genomic DNA or 1-10ng cDNA as a reference. Since the templates of different species contain different copy numbers of the target genes, the templates can be gradient diluted to determine the optimal amount of template to be used.
- (3) The recommended reaction system is 50  $\mu$ l, and the reaction system can be scaled up or down according to the actual experimental needs.
- 2. PCR reaction program:

Caution! The pre-denaturation reaction of this product must be completed at  $95^{\circ}$  C for 10 minutes!

It is recommended to use the two-step PCR shown in the table below to set up the program, and this program is based on the ABI7500 fluorescent quantitative PCR instrument as an example. If you do not get good experimental results due to lower Tm values of primers and other reasons, you can try to carry out three-step PCR amplification, and the three-step procedure is detailed in Optimization of Reaction Conditions.

Step	Temperature	time
Pre denaturation	95° C	10min <sup>1)</sup>
denaturation	95° C	15s 7 25 401
Annealing/Extension <sup>2)</sup>	60° C	$\frac{15s}{1min}$ $35-40$ cycles
Analysis of fusion curve <sup>3)</sup>		
	95° C	15s
	60° C	1min
	95° C	15s
	60° C	15s

Note: 1) The hot-start enzyme used in this product shall be activated by the enzyme under the condition of pre-denaturation  $95^{\circ}$ C and 10min.



- (2) Please use  $60-64^{\circ}\text{C}$  as a reference for setting range of annealing temperature, and increase the annealing temperature when non-specific reaction occurs.
- (3) This program is set up with the ABI7500 Fluorescence PCR instrument as a reference. For melting curve analysis, please set up the program recommended by the fluorescence PCR instrument you are using.

# Optimization of reaction conditions

In the optimization of fluorescence quantification reaction conditions, primer concentration, annealing temperature, and extension time should be considered to improve reaction specificity and amplification efficiency.

- 1. An experimental system with high reaction specificity and amplification efficiency should have the following conditions:
- (1) High reaction specificity: negative control without non-specific amplification such as primer dimer; no amplification beyond the target fragment.
- 2) High amplification efficiency: low Ct value; high PCR amplification efficiency, close to the theoretical value of 100%.
- 2. Methods for optimizing reaction conditions:
- 1) Primer concentration: Usually, a primer concentration of 0.2  $\mu$ M can get better results, and the final concentration of 0.1-1.0  $\mu$ M can be used as a reference for setting the range. If you want to improve the specificity of the reaction, you can reduce the primer concentration; if you want to improve the amplification efficiency, you can increase the primer concentration, thus optimizing the reaction system.
- 2) Annealing temperature: It is recommended to use two-step PCR with an annealing temperature of  $60^{\circ}$  C for the reaction. If you want to improve the specificity of the reaction, you can increase the annealing temperature, and take  $60\text{--}64^{\circ}\text{C}$  as the reference of the setting range. If you can not get good experimental results due to the use of primers with lower Tm values, etc., you can try to perform three-step PCR amplification, and the annealing temperature of the three-step method, please take the range of  $56^{\circ}\text{C}\text{--}64^{\circ}\text{C}$  as a reference for setting.
- 3) Extension time: it is recommended to use two-step PCR with an extension time of 1min for the reaction. If you want to improve the amplification efficiency, you can try to increase the extension time or try three-step PCR. Caution! The pre-denaturation reaction of this product must be completed at 95° C for 10 minutes!

Three-step fluorescence quantitative PCR (this program is based on the ABI7500 fluorescence quantitative PCR instrument):

Step	Temperature	time
Pre denaturation	95° C	10min <sup>1</sup> )
denaturation	95° C	10s
annealing	$56-64^{\circ} \text{ C}^{2}$	$30s$ $32s^3$ ) 35-40 cycles
reach	72° C	$32s^3$ )
Melting curve analysis <sup>3)</sup>		
	95° C	15s
	60° C	1min
	95° C	15s
	60° C	15s



Note: 1) The hot-start enzyme used in this product shall be activated by the enzyme under the condition of pre-denaturation 95°C and 10min.

- 2) Appropriately reduce the annealing temperature when the desired amplification efficiency cannot be obtained; increase the annealing temperature when a non-specific reaction occurs.
- 3) If the reaction amplification efficiency needs to be improved, the extension time can be increased appropriately.
- 4) This program is set up with the ABI 7500 Fluorescence PCR instrument as a reference, for melting curve analysis, please set up the program recommended by the fluorescence quantitative PCR instrument used.